

6β -[2-METHYLBUTANOYLOXY]TROPAN-3 α -OL, A NEW ALKALOID FROM *DATURA CERATOCAULA* STRUCTURE AND BIOSYNTHESIS

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Key Word Index—*Datura ceratocaula*; Solanaceae; tropane alkaloids; 6β -[2-methylbutanoyloxy]tropan-3 α -ol; L-isoleucine-[^3H - ^{14}C] as precursor for 2-methylbutanoyl moiety.

Abstract— 6β -[2-methylbutanoyloxy]tropan-3 α -ol, a new alkaloid, has been isolated from the aerial parts of *Datura ceratocaula* (Solanaceae). Its chemotaxonomic significance within the genus is discussed. Biosynthetically, the 2-methylbutanoyl moiety is derived from L-isoleucine.

THE GENUS *Datura* is divided into four sections, (1) Stramonium, (2) Dutra, (3), Ceratocaulis and (4) Brugmansia. Sections (1) and (2) comprise the more frequently encountered annual herbaceous daturas which have erect flowers and either erect or nodding spiny fruit capsules respectively. The perennial tree-daturas of section (4), unlike the others, readily hybridise but will not self-pollinate and this has caused considerable confusion as to what constitutes a single species. Morphologically, they are distinguished in having long pendulous flowers and large spineless fruits. *Datura ceratocaula*, the only member of section (3), has erect white flowers suffused with lavender and smooth fruits similar to, but smaller than, those of section (4). Because of these similarities and the fact that *D. ceratocaula* will not readily cross with daturas of sections (1) and (2) it has been suggested that it forms a link between section Brugmansia, and sections Stramonium and Dutra.¹⁻³ Geographically, too, its distribution throughout Mexico and Central America, connects through a narrow isthmus the probably more ancient South American Brugmansias with the annual daturas of North America, Europe and Asia. *Datura ceratocaula*, first described by Ortega in 1798, is an aquatic, hollow-stemmed, prostrate, creeping plant long known by the Mexicans as the narcotic "Torna-loca" (maddening plant). It has received little chemical attention, paper chromatography suggesting the presence of hyoscyne, hyoscyamine, cuscohygrine and four unidentified bases.⁴

Analysis of the aerial parts of mature *D. ceratocaula* plants by partition column chromatography at pH 6.8⁵ has now revealed the presence of three bases in the ether eluate.

¹ SAFFORD, W. E. (1920) *Ann. Report Smithsonian Institute Washington*, 537.

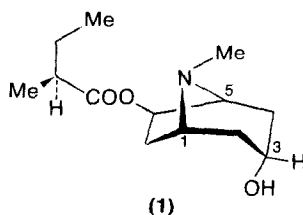
² SAFFORD, W. E. (1921) *J. Washington Acad. Sci.* 173.

³ AVERY, A. G., SATINA, S. and RIETSEMA, J. (1959) in *Blakeslee: The Genus Datura* p. 35 and 220. Ronald, New York.

⁴ ZIELINSKA-SOWICKA, R. and SZLIPCZYNSKA, K. (1968) *Diss. Pharm. Pharmacol.* **20**, 539.

⁵ EVANS, W. C. and PARTRIDGE, M. W. (1952) *J. Pharm. Pharmacol.* **4**, 769.

The third and major base present (0.007%) gave a picrate m.p. 118° and had IR absorption bands at 3420 (OH) and 1740 cm^{-1} (sat. ester carbonyl). As anticipated, the MS was characteristic of a disubstituted tropane⁶ and gave the molecular formula $\text{C}_{13}\text{H}_{23}\text{NO}_2$. The base ion at m/e 113 (100) indicated that the free hydroxyl was at the C(3) position. PMR studies gave the usual pattern of a C(3), (6) disubstituted tropane, the triplet at higher field than usual, $\tau = 5.95$ confirmed that the free hydroxyl was at C(3) and α -orientated.^{7,8} Finally, the characteristic doublet and triplet centred at $\tau = 8.85$ and 9.1 respectively suggested that the C(6) acyl group was 2-methylbutanoic acid and the alkaloid was therefore 6 β -[2-methylbutanoyloxy]tropan-3 α -ol. Hydrolysis of the base gave (+)-2-methylbutanoic acid⁹ and tropan-3 α ,6 β -diol which when esterified with tigloyl chloride afforded (–)-3 α ,6 β -ditigloxytropane, thus confirming the 3R 6R structure assigned.¹⁰ (1) The base was synthesized by esterifying (±)-2-methylbutanoyl chloride with (±)-6 β -hydroxytropan-3-one and reducing the product to the 3 α -ol with Raney nickel.



To date, 2-methylbutanoyl esters have been detected only in sect. *Brugmansia*⁷ and the dehydro derivative of the new base 6 β -tigloyloxytropan-3 α -ol has been isolated from *D. cornigera*,¹¹ and outside the genus in *Anthocersis*,¹² which, like *Duboisia*, is placed in the sub-tribe Salpiglossideae and appears to have an affinity with the tree-daturas. Similarly, 3 α -ol derivatives of di- and tri-hydroxytropane esters have hitherto only been detected in this section. The aerial parts of *D. ceratocaula*, unlike those of the tree-daturas contain little hyoscyne, but the presence of this new ester as a major alkaloid would support Safford's view^{1,2} that there is an affinity between sects. *Brugmansia* and *Ceratocaulis*.

The tiglic acid moiety of the ditigloyl esters of *Datura* is known to be derived from L(+)-isoleucine via 2-methylbutanoic acid (see refs 13 and 14 for scheme) but the biosynthesis of 2-methylbutanoyl esters has not previously been investigated. The isolation of dextrorotatory 2-methylbutanoic acid is in keeping with its derivation from this amino acid, and when L(+)-isoleucine-[U-¹⁴C] was infiltrated into the roots of 6-month-old plants it was incorporated within 2 days. All the radioactivity was located in the 2-methylbutanoyl part.

EXPERIMENTAL

D. ceratocaula plants. Seed obtained from Zentralinstitut für Genetik und Kulturpflanzenforschung, Gatersleben D.D.R. were grown on open land in Leicester and had all the characters previously described.¹⁻³ For analy-

⁶ BLOSSEY, E. C., BUDZIKIEWICZ, H., OHASHI, M., FODOR, G. and DJERASSI, C. (1964) *Tetrahedron* **20**, 585.

⁷ EVANS, W. C. and LAMPARD, J. F. (1972) *Phytochemistry* **11**, 3294.

⁸ PARELLO, J., LONGEVIALLE, P., VETTER, W. and MCCLOSKEY, J. A. (1963) *Bull. Soc. Chim. Fr.* 2787.

⁹ ODHAM, G. (1963) *Ark. Kemi* **20**, 507.

¹⁰ FODOR, G. and SOTI, F. (1965) *J. Chem. Soc.* 6830.

¹¹ EVANS, W. C. and GRIFFIN, W. J. (1963) *J. Chem. Soc.* 4348.

¹² EVANS, W. C. and TREAGUST, P. G. (1973) *Phytochemistry* **12**, 2505.

¹³ BASEY, K. and WOOLLEY, J. G. (1973) *Phytochemistry* **12**, 2197.

¹⁴ BASEY, K. and WOOLLEY, J. G. (1973) *Phytochemistry* **12**, 2883.

sis, the roots and aerial parts were separately dried at 60° for 18 hr. Optical rotations were measured using a Bendix NPL automatic polarimeter type 143. L-isoleucine-[U-¹⁴C] was purchased from the Radiochemical Centre, Amersham.

Counting procedures. Duplicate samples were counted in commercially available toluene or dioxane based POP/POPOP scintillators in a Packard Tri-Carb model 2002 liquid scintillation spectrometer.

Isolation of 6β-[2-methylbutanoyloxy]tropane-3α-ol. In a typical isolation 50 g powdered *D. ceratocaula* "top" was mixed with Ca(OH)₂ (15 g), H₂O (20 ml) and after 30 min exhaustively extracted with cold Et₂O. After distillation of the solvent the green residue was transferred in CHCl₃ (ca 1 ml) to a 10 g kieselguhr column containing 5 ml 0.5 M phosphate buffer pH 6.8. Development with light petrol. (100 ml) removed pigments and Et₂O eluted 3 bases. The second base (0.002%) by titre⁵ was identified as hyoscyne by TLC, Aluminium oxide G (Merck) Et₂O-EtOH 4:1 (*R_f* 0.85) and the third base [α]_D²⁰ + 5.5° (0.007%) (*R_f* 0.8) which was unknown easily gave a picrate (needles) m.p. 118° $\nu_{\text{max}}^{\text{KBr}}$ 3420, 1740, 1635, 1615, 1570, 1550, 1510, 1430, 1365, 1330, 1270, 1140, 1085, 1055, 907, 790, 740 and 705 cm⁻¹; MS (*m/e* with (I) % in parenthesis) 41 (22), 42 (26), 44 (18), 55 (8), 57 (27), 81 (9), 82 (16), 94 (30), 95 (8), 96 (47), 112 (16), 113 (100), 114 (10), 122 (11), 140 (10), 156 (7). *M*⁺ 241 (11). Accurate mass measurement *M*⁺ = 241.1677. C₁₃H₂₃NO₃ requires 241.1678. PMR (Base in CDCl₃ with TMS as internal standard) signals at τ = 4.37 (1H, *dd*, *J* = ca 3.3 cps), 5.9 (1H, *t*, *J* = ca 9 cps), 6.75 (*bm*), 7.5 (3H, *s*), 7.9 (OH, shift with D₂O), 8.85 (3H, *d* *J* = ca 7 cps), 9.1 (3H, *t* *J* = ca 15 cps).

Hydrolysis. The base (from 20 mg picrate) was heated at 100° in a sealed tube with 5% Ba(OH)₂ (10 ml) for 2 hr, when the hydrolysate was acidified (50% H₂SO₄) and extracted with Et₂O (5 × 3 ml). The Et₂O contained (+)-2-methylbutyric acid [α]_D²⁰ + 16.76° (lit.⁹ [α]_D^{21.2} + 19.3°) which was identified by GLC.¹⁵ Excess BaCO₃ was added to the remaining aqueous phase which was then centrifuged and the supernatant liquid was evaporated to dryness under reduced pressure. A mixture of tigloyl chloride (0.4 ml) and the residue (the alkamine sulphate) was heated under reflux for 2 hr,¹⁶ then cooled, acidified and the excess tiglic acid was removed with Et₂O. Extraction of the basified solution with CHCl₃ gave a single base [α]_D²¹ - 18.5° (lit.¹⁷ [α]_D²⁰ - 21.5°), TLC Aluminium oxide G (Merck) Et₂O *R_f* identical with (-)-3α,6β-ditigloyloxytropane, which gave a picrate (4.8 mg, long needles), IR (KBr), m.p., m.m.p. (151°).

Partial synthesis of 6β[2-methylbutanoyloxy]tropan-3α-ol. Dry 6β-hydroxytropan-3-one (28 mg) and 2-methylbutanoyl chloride (0.5 ml) heated under reflux for 2 hr. The recovered base was dissolved in EtOH and hydrogenated (at. press.) in the presence of Raney Ni. The resultant base, picrate mp 124°, had an *R_f* value on TLC and PMR spectrum indistinguishable from the natural base.

Feeding experiments. Thirteen 6-month-old field grown *D. ceratocaula* plants were carefully uprooted, washed free from soil and allowed to stand in blackened beakers containing an aq. soln of L-isoleucine-[U-¹⁴C] (45 μCi) sp. act. 8.7 mCi/mM. After 2 days the roots and aerial parts were separately dried. The aerial parts (60 g) were extracted and the 6β-[2-methylbutanoyloxy]tropan-3α-ol (5.6 mg) from the partition column was diluted by adding authentic base picrate (11 mg) to the shake-out liquors. The re-isolated diluted picrate (9.8 mg) m.p. 118°, sp. act. 5.69 × 10⁴ dpm/mM (calc. sp. incorporation 0.0006%; total incorporation 0.003%) was converted to the base and when hydrolysed gave 2-methylbutanoic acid sp. act. 5.60 × 10⁴ dpm/mM (98.4% recovery of radioactivity) and tropan-3α,6β-diol, isolated as the picrate (3 mg) m.p. 247°.

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¹⁵ BERESFORD P. J. and WOOLLEY, J. G. (1974) *Phytochemistry* **13**, 1249.

¹⁶ BARGER, G., MITCHELL, W. and MARTIN, W. F. (1937) *J. Chem. Soc.* 1820.

¹⁷ EVANS, W. C. and WELLENDOFF, M. (1958) *J. Chem. Soc.* 1991.